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<p>(54) Title: T-CELL LYMPHOTROPHIC VIRUS PROTEIN AND ASSAY</p> <p>(57) Abstract</p> <p>Cells infected with HTLV-III yield a protein (p27) having an apparent molecular weight of about 27,000 daltons. The presence in a biological specimen of p27 antigenic determinants, or antibodies to p27 antigenic determinants is useful information (by itself or as a panel of tests) for predicting the course of HTLV-III infection in seropositive patients. Polypeptides and antibodies used to perform assays to provide that information are disclosed, as well as assays and kits for such assays.</p>			

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T-CELL LYMPHOTROPHIC VIRUS PROTEIN AND ASSAY

Background of the Invention

This invention relates to novel purified forms of a protein found in cells infected with human T-cell lymphotrophic virus III (HTLV-III) and related viruses; the invention also relates to polypeptides, kits, and assays for detecting in a biological specimen the presence of that protein's antigenic determinants or of antibodies to one or more such antigenic determinants. For convenience, the term "HTLV-III related viruses" is used in this application to include viruses that are closely related to HTLV-III by serological biochemical and molecular criteria, including lymphadenopathy associated virus (LAV) (see Barre-Sinoussi (1983) Science 220:859), ARV (see Sanchez-Pescador (1985) Science 227:484) and AAV, and other forms, subtypes and variants, including simian viruses. The terms "human T-cell lymphotrophic virus" and "human T-cell leukemia virus" are used interchangeably, although the former is preferable.

Human T-cell lymphotrophic virus III (HTLV-III) is believed to play a key role in the pathogenesis of acquired immunodeficiency syndrome (AIDS). It has been shown that human patients whose bodies contain antibodies to HTLV-III-infected cells are apparently latently or actively infected with the virus. There are various tests to determine the presence of antibodies to HTLV-III proteins in a biological specimen. For example, an ELISA test is widely used for blood bank screening. For such assays, the goal is to screen out all individuals who have been exposed to HTLV-III, whether or not such individuals have developed or will develop AIDS.

Other HTLV-III-encoded polypeptides that are antigenic when expressed in infected individuals include:

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- 1) a 55 kd gag polyprotein (p55) which yields a 24 kd protein (p24) as the major virus core protein and a 17 kd phosphoprotein (pp17) (Schupbach et al. (1984) Science 224:503-505); and
- 5 2) a gpl60 env polyprotein which gives rise to gpl20 at the amino terminus (see Essex and Lee, USSN 670,361, filed November 9, 1984, which is hereby incorporated by reference).

The disease course for patients who exhibit 10 HTLV-III antibodies varies; some such patients are healthy and currently uninfected, others harbor the virus with few symptoms, still others have chronic symptoms that have been designated "AIDS related complex" (ARC), and, finally, some such individuals are 15 AIDS patients, exhibiting immunodeficiencies that are ultimately fatal. It also appears that some ARC or non-symptomatic patients harboring the virus may develop AIDS after prolonged periods.

Summary of the Invention

20 Cells infected with HTLV-III and related viruses yield a protein coded by the open reading frame (ORF) positioned 3' to the env gene. The protein coded by HTLV-III has an apparent molecular weight of about 27,000 daltons, meaning that, when myristylated, the 25 protein migrates in conventional sodium dodecyl sulfate (SDS) electrophoresis gels as described below in a manner that is characteristic of a molecular weight of about 27,000 daltons, even though the actual molecular weight may differ somewhat from 27 kd; when computed 30 from the viral coding sequence, the HTLV-III protein's molecular weight may be lower, e.g., about 22,000 daltons. For convenience, we refer to the protein coded by the open reading frame 3' to the env gene of HTLV-III and HTLV-III related viruses as "p27". The presence of 35 p27, p27 antigenic determinants, or antibodies to p27 antigenic determinants in patients that are positive for

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HTLV-III or related viruses is useful information (by itself or as part of a panel of tests) for predicting the course of the infection in that patient.

Accordingly, in a first aspect, the invention
5 features substantially pure polypeptides that are useful for assaying for HTLV-III and related viruses antibodies in a human biological specimen. The substantially pure polypeptides contain at least one antigenic determinant that is immunologically cross-reactive with the
10 determinants of a p27 protein obtained from infected cells. By "polypeptides containing immunologically cross-reactive antigenic determinants" is meant polypeptides having in common antigenic determinants with which a given antibody will react.

15 Polypeptides according to the first aspect include the p27 protein itself either myristylated or unmyristylated. If myristylated at the NH₂ terminal, the polypeptide is blocked against Edman degradation. The polypeptides can also include myristylated or
20 unmyristylated fragments of p27. Other useful polypeptides which have the necessary immunogenic determinants include peptides coded by HTLV-III and related viruses as well as synthetic polypeptides, e.g. produced by genetically engineered cells. Preferably,
25 the polypeptide is labeled for use in an assay as described below.

Polypeptides according to the first aspect also include antibodies or fragments thereof which are anti-idiotypic towards the active determinant or
30 determinants on the p27 protein of the invention. It has been shown that anti-idiotypic reagents are useful as diagnostic tools for the detection of antigens carrying sites which are immunologically cross-reactive with those on the antibodies (Potocnjak et al., Science
35 215:1637-1639 (1982)). Thus, an assay for p27

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antibodies could be carried out with the aid of an anti-idiotypic antibody or immunologically active fragment thereof which carries an antigenic site or sites thereon which are immunologically similar to the 5 antigenic site or sites on p27. Such anti-idiotypic antibodies can be raised against primary antibodies having specificity against the antigenic sites on the p27 of the invention (i.e. the anti-idiotypic antibodies are anti-antibodies). Preferably monoclonal 10 anti-idiotypic antibodies are used.

As noted above, the presence of p27 antibodies provides useful information concerning the course of disease in individuals that are positive HTLV-III and related viruses. Accordingly, in a second aspect, the 15 invention features a method of assaying a biological specimen for the presence of p27 antibodies, by incubating the specimen with the above-described polypeptide and determining whether or not an immunocomplex is formed.

In preferred embodiments of the second aspect, the assay for p27 antibodies is performed as part of a panel of tests for HTLV-III and related virus 20 antibodies; thus, one or more specimens from the same patient is assayed for antibodies to other HTLV-III and 25 related virus proteins such as one or more of the following proteins using techniques described in the above-cited references: p55, p24, p17, gp160, gp120, and gp41. The resulting information from positive patients provides an improved basis for estimating the 30 course of the disease. For example, individuals exhibiting either p27 antibodies or p55/p24 antibodies are less likely to have contracted AIDS or ARC than individuals who are negative for both p55 and p24 35 antibodies; and those who are positive for both antibodies are even less likely to have contracted AIDS or ARC.

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The invention also features assaying for the presence of a p27 antigenic determinant or for determinants cross-reactive with the determinants of p27. The determinants to be assayed may occur on p27 itself or on other polypeptides. The determinants may be in free circulation in body fluids or in lymphocytes. The assay can be carried out by known immunoassay methods, using antibodies, monoclonal or polyvalent, having immune reactivity with the antigenic determinants found on p27. For example competitive immunoassays or immunometric (sandwich) assays can be used.

Finally, the invention features antibodies that are useful in the above-described assays because they have antigenic determinants which are immunologically reactive with p27.

Other features and advantages of the invention will be apparent from the following description of the preferred embodiment and from the claims.

20 Description of the Preferred Embodiment
Drawings

Fig. 1 is a diagrammatic representation of HTLV-III DNA indicating coding regions of various proteins.

Fig. 2 is a graph depicting radioactivity detected in peptide fragments of p27 and depicting the amino terminal amino acid sequence of two such peptide fragments.

Fig. 3 is a photograph of SDS-polyacrylamide gels of radioimmunoprecipitates of HTLV-III infected cell proteins.

The Protein

The protein of the present invention, when myristylated, has a molecular weight of approximately 27,000 daltons as determined by sodium dodecyl sulfate

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(SDS) gel electrophoresis and is soluble in SDS buffer consisting of 0.15 M sodium chloride, 0.05 M Tris hydrochloride pH 7.2, 1% Triton X-100, 1% sodium deoxycholate, 0.1% sodium dodecylsulfate, and 1 mM phenylmethylsulfonyl fluoride. Triton X-100 is a nonionic detergent (octylphenoxy polyethoxy (9-10) ethanol). The p27 protein undergoes post-translational myristylation, and is not glycosylated. The unmyristylated moiety of the protein contains substantially the same antigenic determinant or determinants as does the myristylated protein. An NH₂-terminal myristylate group blocks Edman degradation.

Further characterization of the p27 protein, and indication that it is distinct from other, previously described HTLV-III specific proteins listed above, can be obtained by mapping the protein to HTLV-III DNA. p27 protein is obtained as described below from HTLV-III infected cells labeled with [³H] leucine and [³H] valine. The protein is fragmented with cyanogen bromide, and the fragments are separated by Sephadex S-200 chromatography. Fractions are analyzed by Edman degradation and pooled.

Fig. 2 shows a diagrammatic representation of four peptides that are predicted to be generated by cyanogen bromide cleavage at met residues at positions 1, 20, 60, and 170 of the 204 amino acid sequence deduced from the 3' ORF of HTLV-III DNA. To match p27 to that HTLV-III DNA coding region, H9/HTLV-III_{E-84-4} cells (described below) are metabolically labeled with 10mCi of [³H] leucine or [³H] valine for four hours and cell lysates prepared. Radiolabeled lysates are precipitated with standard HTLV-III reference serum (same as Fig. 1, lane 2) and electrophoresed on 10% SDS-polyacrylamide gels. The p27

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is excised from the gel, electroeluted and lyophilized prior to cyanogen bromide cleavage. Cyanogen bromide hydrolysis of p27 is accomplished by incubating the protein for nine hours with two percent CNBr in seventy percent formic acid. The reaction is terminated with the addition of nine volumes of water and the mixture is lyophilized. Sephadryl S-200 separation of CNBr peptides and automated sequence analysis are performed by the general technique of Lee et al., Science (1984) 226:57 et seq. and Coligan et al. Methods Enzymol. (1983) 91:413. Fractions are collected, assayed for radioactivity, pooled, and subjected to Edman degradation. The sequences derived from pools C and F are compared to the amino acid sequences of CNBr cleavage products of the gene product coded by the 3' open reading frame (ORF) of HTLV-III derived from the DNA sequence data given by Meusing et al. (1985) Nature (London) 313:459 or Sanchez-Pescador (1985) Science 227:484.

The amino terminal sequence data derived from Edman degradation of peptides actually derived from pools C and F matches the predicted sequences shown in Fig. 2. Accordingly, the coding region for p27 is the open reading-frame (ORF) at the 3' terminal region of HTLV-III. The labeled amino acids are indicated by asterisks. It is likely that the methionine initiator sequence for the p27 coding region is the AUG codon in the 3' ORF which begins 5 nucleotides from the 3' end of the env gene and is followed by a glycine codon.

Fig. 3 is a photograph of an SDS-Page radioimmunoprecipitation of HTLV-III proteins showing antibody reactivities specific to p27. Cell lysates are prepared from [³⁵S] cysteine labeled H9/HTLV-III E-84-4 cells or H9 uninfected cells.

Radioimmunoprecipitation and SDS-PAGE are carried out by

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the method generally described in Kitchen et al. (1984) Nature 312:367; Barin et al. (1985) Science 228:1094; and Allan et al. (1985) Science 228:1091. Lysates are immunoprecipitated with the following sera: normal human
5 serum pre-tested on uninfected H9 cells (lane a) and H9/HTLV-III_{E-84-4} cells (lane 1); positive reference serum from an ARC patient pre-tested on uninfected (lane b) and infected cells (lane 2); serum from seven ARC patients (lanes 3, 4, 5, 8, 9, 10, and 13); serum from 10 three AIDS patients (lanes 6, 11, and 12); and serum from one healthy homosexual male (lane 7). The serum of lanes 3-12 is tested on infected H9/HTLV-III_{E-84-4} cells. Immunoprecipitates are electrophoresed on 20 centimeter 10% SDS-polyacrylamide gels.

15 Obtaining the Protein

The protein can be obtained from HTLV-infected cells. A variety of cell lines have been prepared, which are permanently and persistently infected with HTLV; among them can be mentioned HTLV-III-infected H9 cells, Lymphadenopathy associated virus (LAV)-infected NC37 cells, and Molt 3 and HUT78 cells infected with fresh AIDS virus isolates. One particular such HTLV-III infected cell line is H9G/HTLV-III_{E-84-4} (ATCC 8983), an isolate produced by repeated passage of 20 H9/HTLV-III (ATCC 8543). It may be that the exact sizes of the novel protein is slightly different in different lines; however, the common immunologically cross-reactive portion of the protein is the same regardless of cell line, since it is a protein induced 25 by HTLV. Thus, any cell which harbors the virus may be an appropriate source for the novel proteins.

In order to obtain the protein from any infected cells carrying the virus, the cells are metabolically labeled (e.g. with ³⁵S-cysteine) and

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immunoprecipitated with antisera obtained from HTLV-III-infected subjects. The novel proteins can then be detected and isolated in the cell lysate by SDS gel electrophoresis.

5 A specific protocol of p27 preparation is as follows. Infected HTLV-III cells are harvested at their log phase of growth. After one wash with RPMI-1640 medium, each sample of the cells is resuspended in a labeling medium consisting of RPMI-1640 medium, 10%
10 fetal bovine serum, and 100 μ Ci/ml of 35 S-methionine. At the end of about 4 to 8 hours pulsing, the radioactive labeled cells are washed three times with cold PBS. The cell pellet is then lysed with 0.2 ml/1 $\times 10^6$ cells of cold lysis buffer (RIPA) (0.15
15 M sodium chloride, 0.05 M Tris-hydrochloride pH 7.2, 1% Triton X-100 wetting agent, 1% sodium deoxycholate, 0.1% sodium dodecyl sulfate, and 1 mM phenylmethylsulfonyl fluoride). After 10 minutes of intermittent vortexing, the mix is centrifuged for 1 hour at 100,000 xg at 4°C
20 and absorbed on Sepharose Cl-4B-protein-A beads. The lysate is then subjected to immunoprecipitation with positive anti-sera, and the immunoprecipitate is electrophoresed.

25 An alternative method for preparing the protein, the 3' ORF HTLV-III region can be isolated and cloned for expression in a bacterial expression system such generally using the technique described in Chang et al. (1985) Nature 315:151.

Assays

30 The purified and isolated proteins or any polypeptide antigen immunologically cross-reactive therewith can be employed as a standard antigen in any conventional assay procedure for detection in biological specimens of the presence of antibodies specific
35 thereto.

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The proteins or polypeptides immunologically cross-reactive therewith can be labeled by conventional procedures with ^{125}I or ^{35}S or ^3H for use in radioimmunoassay, with fluorescein for fluorescent immunoassay, with enzyme for enzyme immunoassay or with biotin, for biotin-avidin linked assays. It can be employed, labeled or unlabeled as desired, in competitive immunoassays, as well as in double antibody assays using two antibodies, either of the 5 idiotype:anti-idiotype variety or more particularly of the second antibody type using an anti-Fc antibody, or other assays.

To form the protein antibody complex, aliquots of protein-A-coated beads are bound to: (a) positive 15 reference blood serum from an individual known to harbor antibodies against HTLV-III viral proteins; (b) negative control serum from individuals free from infection; and (c) serum from unknown individuals to be tested. Each aliquot of coated beads is then reacted with an aliquot 20 of each precleared lysate obtained from cells according to the methods described above at 4°C for 4-8 hours to permit complex formation or immunoprecipitation to occur between the bonded lysate protein and any antibody present in the sera. At the end of the reaction the 25 beads are washed 5 times with the buffer (RIPA) to remove uncomplexed lysate protein.

The beads are then immersed in a sample buffer (0.1 M Cleland's reagent, 2% sodium dodecylsulfate, 0.08 M Tris-hydrochloride pH 6.8, 10% glycerol, and 0.2% 30 Bromphenol Blue) and subjected to boiling at 100°C for 2 minutes to elute proteins from the beads and to dissociate complexes.

Alternatively, the novel proteins or polypeptides immunologically cross-reactive therewith 35 could be immobilized on an insoluble phase, such as an

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insoluble resin, and detection of the anti-protein antibodies can be carried out by measuring their binding to the insoluble phase.

Insoluble phases also include latex particles, which

5 when coated with the novel protein or its immunologically cross-reactive polypeptides and subjected to anti-protein antibody, will agglutinate. Yet other insoluble phases include test tubes, vials, titration wells, and the like, to which the novel

10 protein or its immunologically cross-reactive polypeptide can be bound, and antibody thereto detected by double antibody techniques or Protein-A dependent techniques.

The assay for p27 antibody may utilize the p27 protein in crude form and is not limited to using this

15 protein in substantially pure form. For example, the protein may be substantially purified, or cruder mixtures can be used.

Preferably, the assay for p27 or p27 antibodies is performed on individuals who are seropositive, that

20 is, who demonstrate the presence of antibodies to one of the relatively strongly antigenic glycoproteins (gp120, gp160, and gp41). The p27 antibody assay provides the following information:

25 Disease Course for HTLV-III (gp120)
Seropositive Individuals

	<u>Symptom-free</u>	<u>ARC</u>	<u>AIDS</u>	<u>Total</u>
anti-p27+	48.1%	42.9%	29.0%	37.0

Also, preferably, knowledge that an HTLV-III positive

30 patient has anti-p27 antibodies is combined with information gained from other tests to increase the reliability of an assessment of the course of the virus in that patient. For example, the presence of antibodies to other weakly antigenic HTLV-III or related

virus proteins (e.g., p55 or p24) is further information to be used with the presence of anti-p27 antibodies. Such tests can be used as a panel to predict the disease course.

5 Preferably, the tests for the various HTLV-III antibodies are performed by the methods indicated in the following table:

Antigen	Genomic Origin in Virus	Richest Source*	Optimal Procedures for Antibody Testing*
10. p55	<u>gag</u> (entire)	IC*	WBIC, IPIC, CIF*
p24:	<u>gag</u> (middle)	CV	E, WBCV, WBIC, ICV, IPIC, CIF
p27	3' <u>orf</u> (entire)	IC	IPIC

*Abbreviations: CIF, cytoplasmic immunofluorescence; CV, concentrated virus; E, ELISA; IC, infected cell; 15 ICV, immunoprecipitation with concentrated virus; IPIC, immunoprecipitation with infected cell homogenate; WBIC, Western Blotting with infected cell homogenate; WBCV, Western blotting with concrated virus.

The elements necessary for carrying out the 20 diagnostic methodology described hereinbefore may be present in a kit. Such kit comprises a carrier being compartmentalized to receive therein one or more containers, each of said containers comprising one or more elements necessary to carry out the tests.

25 For example, the first container may contain one or both of the purified protein p27 or its immunologically cross-reactive polypeptides in detectably labeled or in insolubilized form.

A second container may comprise anti IgG 30 antibody, polyclonal or monoclonal, useful in double antibody binding assay, or elements needed for detection

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of the label on the protein or its immunologically cross-reactive polypeptides (e.g. chromogenic substrates).

Additional containers may comprise varying 5 amounts of the protein or its immunologically cross-reactive polypeptides which can be used to prepare a standard curve into which experimental results can be interpolated. The materials may be present in the kit by themselves, in solution, freeze-dried, or in 10 admixture with other inert materials, such as inert proteins, and the like.

The biological specimens tested may include blood, serum, lymphocytes, urine, tissues, saliva, feces, and the like. Other protein antibodies may be 15 added to the test panel, for example, antibodies to proteins coded by viruses that are closely related to HTLV-III.

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Claims

1 1. The substantially pure polypeptide having
2 at least one antigenic determinant or determinants
3 immunologically cross-reactive with a determinant of a
4 protein coded by the open reading frame 3' to the env
5 gene of HTLV-III or a related virus.

1 2. The polypeptide of claim 1 wherein said
2 protein has an apparent molecular weight of
3 approximately 27,000 daltons.

1 3. The polypeptide according to claim 1
2 wherein said polypeptide is unmyristylated, and
3 comprises to said protein or a fragment of said protein.

1 4. The polypeptide according to claim 1
2 wherein said polypeptide is myristylated and comprises
3 said protein or a myristylated fragment of it.

1 5. The polypeptide according to claim 1
2 wherein said protein is myristylated.

1 6. The polypeptide according to claim 1
2 wherein said protein is inaccessible to Edman
3 degradation from the amino terminus.

1 7. The polypeptide according to claim 1
2 wherein said protein comprises a segment of at least six
3 amino acids from the sequence:
4 -HN-Asp-Asp-Pro-Glu-Arg-Glu-Val-Leu-Glu-Trp-Arg-
5 Phe-Asp-Ser-Arg-Leu-Ala-Phe-His-His-Val-Ala-Arg-Glu-Leu-
6 His-Pro-Glu-Try-Phe-Lys-Asn-Cys-COO-.

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1 8. The polypeptide according to claim 1 or
2 claim 6 wherein said protein comprises a segment of at
3 least six amino acids from the sequence:
4 -HN-Arg-Arg-Ala-Glu-Pro-Ala-Ala-Asp-
5 Gly-Val-Gly-Ala-Ala-Ser-Arg-Asp-Leu-Glu-Lys-His-Gly
6 Ala-Ile-Thr-Ser-Ser-Asn-Thr-Ala-Ala-Thr-Asn-Ala-Ala-
7 Cys-Ala-Trp-Leu-Glu-Ala-COO.

1 9. The polypeptide of claim 1 wherein said
2 protein is coded by the 3' ORF DNA of HTLV-III from
3 ATCC 8943, or by DNA hybridizable to said 3' ORF DNA.

1 10. The polypeptide of claim 1 wherein said
2 protein has a calculated molecular weight of about 22 kd
3 based on the DNA sequence encoding said protein.

1 11. The polypeptide according to claim 1
2 wherein said polypeptide comprises an antiidiotypic
3 antibody having antigenic determinants which are
4 cross-reactive with those of said protein.

1 12. The polypeptide according to claim 1
2 wherein said polypeptide is detectably labeled.

1 13. The polypeptide according to claim 1
2 wherein said polypeptide is bound to an insoluble phase.

1 14. An antibody having at least one antigenic
2 determinant that is immunologically cross-reactive with
3 said polypeptide of claim 1 and said protein of claim 1.

1 15. The method of assaying a biological
2 specimen for the presence of antibody to cells infected
3 with HTLV-III or a related virus, said method comprising

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4 incubating said specimen with a polypeptide
5 having an antigenic determinant or determinants
6 immunologically cross-reactive with determinants of a
7 protein having an apparent molecular weight of
8 approximately 27,000 daltons, said protein being
9 myristylated and being obtained from the cells infected
10 with human T-cell leukemia virus or a related virus, and
11 determining whether or not an immunocomplex is
12 formed between said antibody and said polypeptide.

1 16. The method according to claim 15 including
2 the step of labeling said polypeptide.

1 17. The method of claim 15 further comprising
2 assaying said biological specimen for the presence of a
3 second antibody to cells infected with HTLV-III or a
4 related virus by incubating said specimen with a second
5 polypeptide and determining whether or not an
6 immunocomplex is formed between said second antibody and
7 said second polypeptide, said second polypeptide having
8 an antigenic determinant immunologically cross-reactive
9 with determinants of a second protein obtained from the
10 cell surface of cells infected with HTLV-III or a
11 related virus, said second polypeptide lacking antigenic
12 determinants immunologically cross-reactive with said
13 first protein or said first polypeptide.

1 18. The method of claim 17 wherein said second
2 protein is a gag protein or an env protein.

1 19. The method of claim 17 wherein said second
2 protein is selected from p55, p24, pp17, gp160, gp120,
3 p55, and gp41.

1 20. The method of claim 19 wherein said second
2 protein is p55.

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1 21. A method of detecting the presence in a
2 biological specimen of an antigenic determinant or
3 determinants immunologically cross-reactive with those
4 of the polypeptide claimed in claim 1 which comprises:
5 incubating said specimen with antibodies
6 against the polypeptide of claim 1, and
7 determining whether or not an immunocomplex is
8 formed between said antibodies and said polypeptides.

1 22. The method as claimed in claim 21 wherein
2 said specimen comprises human lymphocytes.

1 23. A kit useful for assaying a sample for the
2 presence of antibody to cells infected with HTLV-III or
3 a related virus, said kit being compartmentalized to
4 receive in close confinement therein one or more
5 containers which comprises
6 a first container containing the polypeptide of
7 claim 1, and
8 a second container containing means for
9 detecting the formation of an immunocomplex between said
10 antibody and said polypeptide.

1 / 3

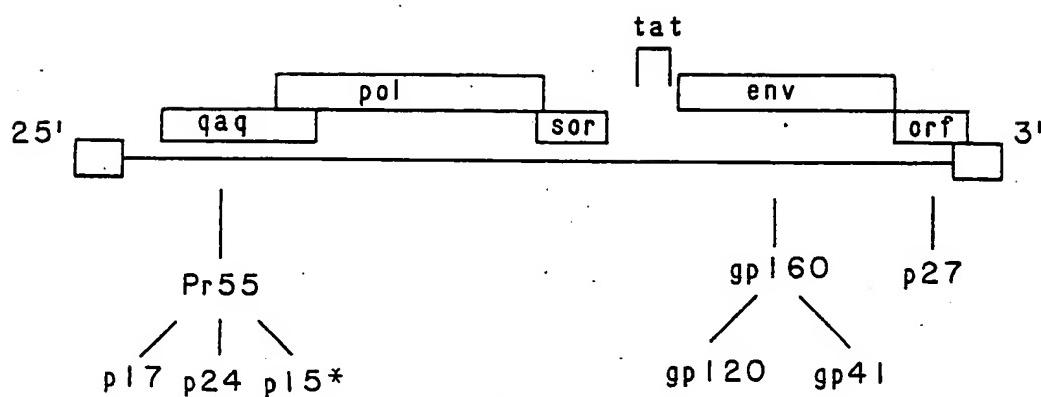


FIG. 1

2 / 3

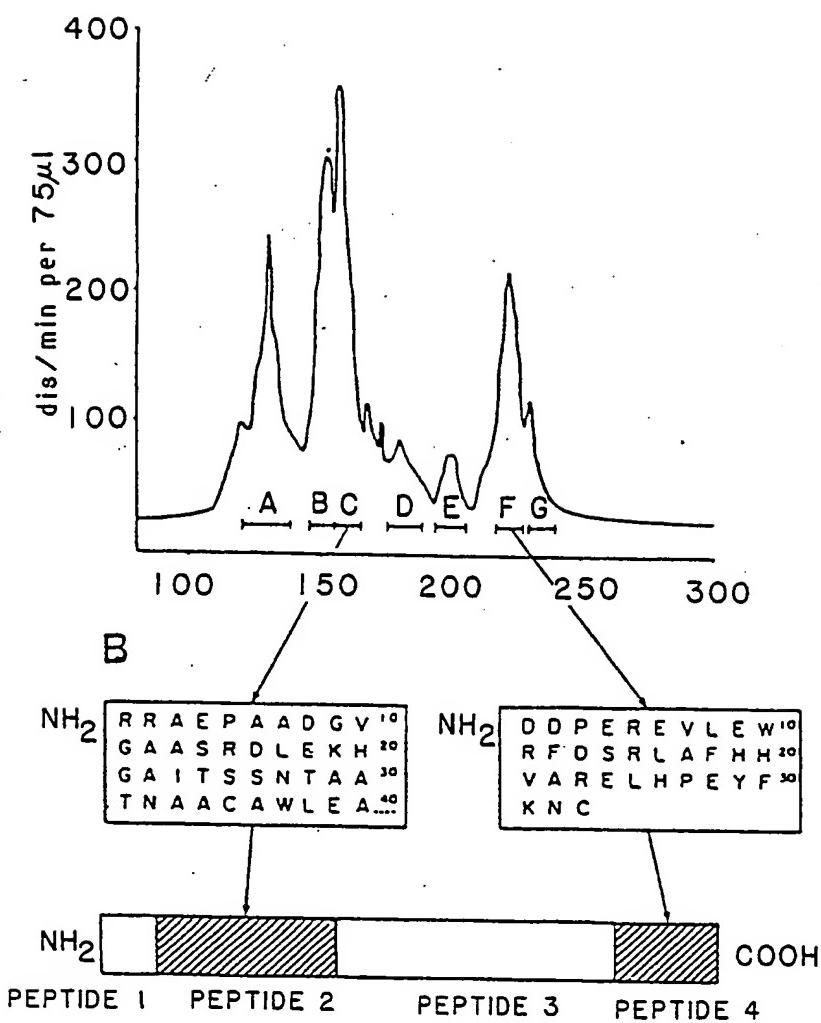


FIG. 2

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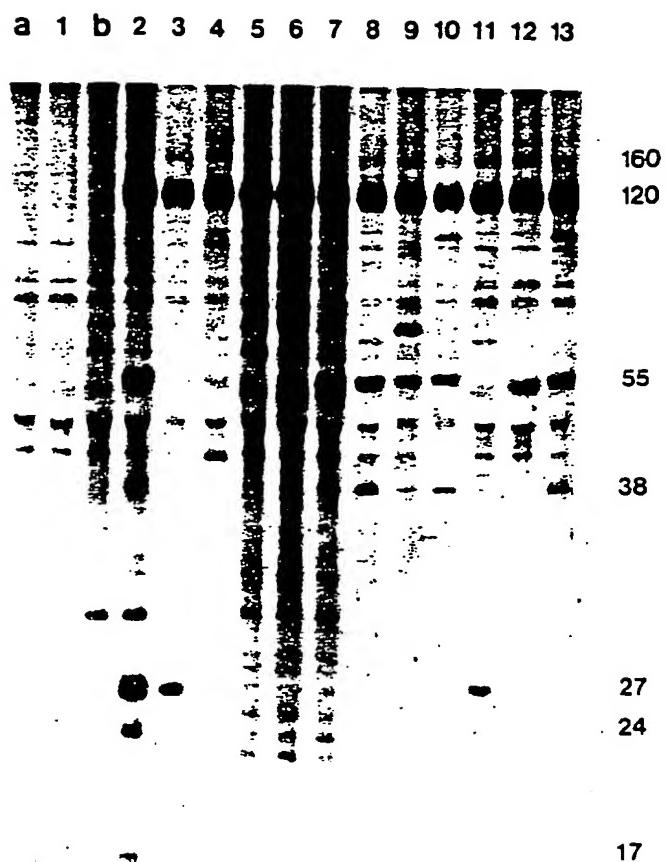


FIG. 3

SUBSTITUTE SHEET

INTERNATIONAL SEARCH REPORT

International Application No PCT/US86/02381

I. CLASSIFICATION OF SUBJECT MATTER (If several classification symbols apply, indicate all) ¹³

According to International Patent Classification (IPC) or to both National Classification and IPC
 U.S. CL. 530/324, 530,387; 435/5,7,810; 436/501,810
 INT. CL. (4) C07K 7/10,15/00; C12Q 1/70; . . .
 G01N 33/566

II. FIELDS SEARCHED

Minimum Documentation Searched ¹⁴	
Classification System	Classification Symbols
US	530/324,530,387; 435/5,7,810 436/501,810
Documentation Searched other than Minimum Documentation to the Extent that such Documents are Included in the Fields Searched ⁵	

III. DOCUMENTS CONSIDERED TO BE RELEVANT ¹⁴

Category ⁶	Citation of Document, ¹⁶ with indication, where appropriate, of the relevant passages ¹⁷	Relevant to Claim No. ¹⁸
A,	PROCEEDING NATIONAL ACADEMY SCIENCE USA, VOL. 81, pages 6202-6206, ISSUED OCTOBER 1984, (Kiyokawa)	1-23
P,A,	PROCEEDING NATIONAL ACADEMY SCIENCE ISSUED DECEMBER 1985, VOL. 82, pages 8359-8363, (Kiyokawa)	1-23
P,A,	CHEMICAL ABSTRACT, VOL. 104, ISSUED 1986, abstract no. 182928e, (Markham)	1-23
P,X,	CHEMICAL ABSTRACT, VOL. 104, ISSUED 1986, abstract no. 1644r, (Chang)	13-23
Y,	CHEMICAL ABSTRACT, VOL. 103, ISSUED 1985, abstract no. 52370k, (Schneider)	1-12
A,	CELL, VOL. 41, pages 979-986, ISSUED JULY 1985, (Crowl)	1-23

* Special categories of cited documents: ¹⁵

- "A" document defining the general state of the art which is not considered to be of particular relevance
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"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"&" document member of the same patent family

IV. CERTIFICATION

Date of the Actual Completion of the International Search ²

16 JANUARY 1987

Date of Mailing of this International Search Report ³

02 FEB 1987

International Searching Authority ⁴

ISA/US

Signature of Authorized Officer ¹⁰

III. DOCUMENTS CONSIDERED TO BE RELEVANT (CONTINUED FROM THE SECOND SHEET)		
Category *	Citation of Document, ¹⁶ with indication, where appropriate, of the relevant passages ¹⁷	Relevant to Claim No ¹⁸
X,	NATURE, VOL. 315, ISSUED 9 MAY 1985 pages 151-154, (Chang).	1
Y,	NATURE, VOL. 313, ISSUED 24 JANUARY 1985, pages 277-284, (Ratner)	1-12
A,	VIROLOGY, VOL. 136, ISSUED 1984, pages 338-347, (Hattori)	1-23
Y,	PROCEEDING NATIONAL ACADEMY SCIENCE VOL. 78, No. 6, pages 3824- 3828, ISSUED JUNE 1981, (Hopp)	1-12
Y,	SCIENCE, VOL. 228, ISSUED 5 APRIL 1985 pages 93-96, (Chang)	1-12
X,	CHEMICAL ABSTRACT, VOL. 102, ISSUED 1985, abstract no. 42543j, (Koyanagi)	1-23
P,X,	CHEMICAL ABSTRACT, VOL. 104, ISSUED 1986, abstract no. 162751p (Lee)	1-13
P,X,	CHEMICAL ABSTRACT, VOL. 104, ISSUED 1986, abstract no. 162752q, (Sodroski)	1-13
P,X,	CHEMICAL ABSTRACT, VOL. 104, ISSUED 1986, abstract no. 31295v, (Pauletti)	1-12
A,	SCIENCE VOL. 224, ISSUED 4 MAY 1984 pages 503-505, (Schupbach)	1-23
X,	SCIENCE, VOL. 215, ISSUED 26 MARCH 1982 No. 4540, pages 1637-1639, (Potocnjak)	1-23
A	SCIENCE, VOL. 22, ISSUED 4 MAY 1984 No. 4648, pages 506-508, (Sarngadharan)	1-23
X	NATURE, VOL.312, ISSUED 22 NOVEMBER 1984, pages 367-369 (Kitchen)	13-23
Y,	SCIENCE, VOL. 228, No. 4699 ISSUED 3 MAY 1985, pages 593-595 (Robey)	1-12
A,	PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES, VOL. 82, pages 3481-3484 ISSUED MAY 1985 , (Sarngadharan)	1-23

FURTHER INFORMATION CONTINUED FROM THE SECOND SHEET

Y,	SCIENCE, VOL. 228, no. 4703, ISSUED 31 MAY 1985, pages 1091-1094, (Allan)	l-12
Y	SCIENCE, VOL. 228, ISSUED 31, MAY 1985, pages 1094-1096, (Barin)	l-12
P, Y, US, A,	4,554,101, published 19 Nov. 1985, (Hopp)	l-12
X, US, A,	4,520,113, Published 28 May 1985 (Gallo)	13-23

V. OBSERVATIONS WHERE CERTAIN CLAIMS WERE FOUND UNSEARCHABLE¹⁰

This international search report has not been established in respect of certain claims under Article 17(2) (a) for the following reasons:

1. Claim numbers because they relate to subject matter¹¹ not required to be searched by this Authority, namely:

2. Claim numbers because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out¹², specifically:

VI. X OBSERVATIONS WHERE UNITY OF INVENTION IS LACKING¹³

This International Searching Authority found multiple inventions in this international application as follows:

I. Claims 1-13 drawn to synthetic peptides classified in Class 530 Subclass 324 and 530.

II. Claim 14 drawn to antibodies classified in Class 530 Subclass 387.

As all required additional search fees were timely paid by the applicant, this International search report covers all searchable claims of the international application.

As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims of the international application for which fees were paid, specifically claims:

No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claim numbers:

As all searchable claims could be searched without effort justifying an additional fee, the International Searching Authority did not invite payment of any additional fee.

Remark on Protest

The additional search fees were accompanied by applicant's protest.

No protest accompanied the payment of additional search fees.

PCT/US86/02381

III. Claims 15-23 drawn to test methods and Kits classified
in Class 435 Subclass 5, 7 and 810 also Class 436
Subclass 501 and 810.